

RESEARCH ARTICLE

Phylogeny and New Classification of Hydrothermal Vent and Seep Shrimps of the Family Alvinocarididae (Decapoda)

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Abstract

The paper addresses the phylogeny and classification of the hydrothermal vent shrimp family Alvinocarididae. Two morphological cladistic analyses were carried out, which use all 31 recognized species of Alvinocarididae as terminal taxa. As outgroups, two species were included, both representing major caridean clades: Acanthephyra purpurea (Acanthephyridae) and Alpheus echiurophilus (Alpheidae). For additional support of the clades we utilised available data on mitochondrial Cytochrome c Oxidase I gene (CO1) and 16S ribosomal markers. Both morphological and molecular methods resulted in similar tree topologies and nearly identical clades. We consider these clades as evolutionary units and thus erect two new subfamilies: Rimicaridinae (Alvinocaridinides, Manuscaris, Opaepele, Shinkaicaris, Rimicaris), Alvinocaridinae (Alvinocaris), whilst recognising Mirocaridinae (with genera Mirocaris and Nautilocaris) at subfamily level. One genus, Keldyshicaris could not be assigned to any subfamily and is thus left as incertae sedis. The monophyly of Alvinocardinae was supported by morphological data, but not supported by molecular data (two analyses); the monophyly of all subfamilies was supported both by morphological and molecular data. Chorocaris is herein synonymized with Rimicaris, whilst Opaepele vavilovi is herein transferred to a new genus Keldyshicaris. Morphological trends within Alvinocarididae are discussed and short biogeographical remarks are given. We provide emended diagnoses for all subfamilies and genera along with keys to all recognized species.

Introduction

Shrimps of the family Alvinocarididae inhabit deep-sea cold-seeps and hydrothermal vent areas around the world, and have been found in the Atlantic, Pacific, and Indian Oceans [1] within the depth range of 252 to 4960 m [2–3]. Most species of the family occur at hydrothermal vents, but a few are found in cold-seep areas [3]; one species, *Alvinocaris longirostris*, has been reported from both vents and seeps [4–7]. The first record of the family was based on a few specimens from the Galapagos Rift, which were described in 1982 as *Alvinocaris lusca* by Austin Williams and Fenner Chace [8]. Later the first author described a further two species of



a new genus, *Rimicaris* from the hydrothermal vent field TAG [9]. One of these species was subsequently transferred to a new genus *Chorocaris* in 1990 [10]. In the middle of the 1990s, Russian and American scientists described two further genera *Opaepele* [11] and *Mirocaris* [12]; whilst more recently the genera *Nautilocaris*, *Shinkaicaris*, and *Alvinocaridinides* were described by Japanese and French researchers [2], [13], [14].

Due to drastic metamorphosis in ontogeny, the history of the family systematics has not been smooth and some taxa were later synonimised. For example, the genus *Iorania* [15] and the species *Rimicaris aurantiaca* [16] are now considered to be juveniles of *Rimicaris exoculata*. There was no consensus on the status of the new family Mirocarididae established for a single genus *Mirocaris* [12]; phylogenetics showed a significant distance between this group and the rest of Alvinocarididae [17–18], although taxonomists kept *Mirocaris* as a genus within Alvinocarididae [19]. Status for *Opaepele vavilovi* also remains unseratin [20].

Three new species and a new genus *Manuscaris* have recently been described from hydrothermal vents in the Pacific Ocean [21]. In this comprehensive study, partial sequences of mitochondrial COI were used, resulting in a minor change in the classification of Alvinocarididae, the transfer of *Opaepele susannae* into *Chorocaris* [21].

At present, 9 genera and 31 species are known within the family and a comprehensive phylogenetic analysis is needed to disentangle existing problems and to elucidate the status of all genera.

This task is eased by the presence of a significant amount of information on partial sequences of mitochondrial COI gene in GenBank, unusually rich for decapods. Much of this data was used in a previous comprehensive study [17], which confirmed three distinct clades consistent with morphology at that time: (1) *Rimicaris/Chorocaris/Opaepele*, (2) *Alvinocaris*, and (3) *Mirocaris*. Evolutionary relationships of vent-endemic shrimp species were shown to correlate neither with their current biogeographic distribution nor with the history of sea-floor spreading. Later studies have incorporated further molecular data for several recently described species and enhanced information for the species studied in [3], [17], [18], [21–26]. However, no attempt to carry out a complete phylogenetic analysis of the whole family Alvinocarididae has been carried out to date.

Combining both morphological and molecular evidence should shed light on the complex relationships in Alvinocarididae.

In this paper we summarize original and literature data about the composition, morphology, and genetic diversity of the family Alvinocarididae. Further, we (1) find and describe morphological characters, (2) perform cladistic morphological analyses, (3) analyze molecular data, (4) combine and compare morphological and molecular results, (5) discuss supported taxa, and (6) provide a new classification, emended diagnoses, and identification keys for all subfamilies, genera, and species.

Material and Methods

Material for morphological analysis

Material was collected along the Mid-Atlantic Ridge during six cruises of R/V "Akademik Mstislav Keldysh" with the use of two deep-sea manned submersibles "Mir-1" and "Mir-2" (34th cruise, August-October 1994, 39th cruise, August-October 1996, 41st cruise, August-December 1998, 47th cruise, June-July 2002, 49th cruise August 2003, 50th cruise, August 2005). Seven vent fields were investigated during 1994–2005, including Menez Gwen (37.8417 N 31.525 W), Lucky Strike (37.2933 N 32.2733 W), Rainbow (36.23 N 33.902 W), Broken Spur (29.17 N 43.1717 W), TAG (26.1367 N 44.8267 W), Snake Pit (23.3683 N 44.95 W) and Logatchev



(14.752 N 44.9785 W). No specific permission was required for field studies in any of these locations. The field studies did not involve endangered or protected species.

Shrimps were collected using baited traps and suction samplers. Immediately after retrieval all specimens were sorted, measured, and preserved in 80% alcohol. Measurements follow established methods for shrimp morphological description [27]. Shrimp morphology and its temporal/spatial variations were thoroughly investigated for this material on the basis of 5861 individuals [28], [29]. A detailed description of this material and discussion of the various species may be found in [20], [29], [30].

Analysis of the morphology of all species within the family was made with the use the above original data and all other available literature data (<u>Table 1</u>).

Terminal taxa, outgroups, and characters used for morphological analysis

All thirty-one recognized species of Alvinocarididae were included as terminals. Outgroup selection was made on the basis of a comprehensive molecular study [31], which revealed two major clades of Caridea: (1) Alpheidae, Hippolytidae, Crangonidae, Glyphocrangonidae, Barbouriidae, Pandalidae, Hymenoceridae, Gnathophyllidae, and Palaemonidae and (2) Rhynchocinetidae, Oplophoridae, Nematocarcinidae, Alvinocarididae, Campylonotidae, Pasiphaeidae and Eugonatonotidae. The first outgroup species, *Acanthephyra purpurea* A. Milne-Edwards, 1881 [32], represents the first clade: Wong et al. [33] have shown that family Acanthephyridae is sister to Oplophoridae and advocate combining both families as Oplophoridae. We chose *A. purpurea* partly because this species is present in GenBank and could also be used as the outgroup in the molecular analysis. The second outgroup species, *Alpheus echiurophilus* Anker, Komai and Marin 2015 [34], belongs to Alpheidae and represents the second major clade of Caridea. Both species are ecologically very different (pelagic and burrowing) as well as morphologically and a comparison of cladograms is thus instructive.

Sixty-three morphological characters (ten multistate) were used in the analysis, and are listed in <u>Table 2</u>, along with character states, brief descriptions, and references to figures (see also Figs 1-3). The data matrix is presented in <u>Table 3</u>.

Analytical method for cladistic analysis

Data were analyzed using a combination of programs by maximum parsimony: Winclada/Nona, TNT, and Mesquite [35–37].

All characters were unordered (non-additive) and equally weighted, missing data were scored as unknown. Characters were unordered, so the score given for each state (i.e., 0, 1, 2) implies nothing about order in a transformation series [38]. Trees were generated in TNT under the implicit enumeration. Relative stability of clades was assessed by standard bootstrapping (sample with replacement) with 10000 pseudoreplicates and by Bremer support (algorithm TBR, saving up to 10000 trees up to 3 steps longer).

Molecular data

Both Mitochondrial Cytochrome c Oxidase I (CO1) and 16S ribosomal markers were selected for phylogenetic analyses, as only these markers have been sequenced for a representative number of alvinocaridid species, with CO1 sequences for 20 (out of 31) species available (<u>Table 4</u>). For the present phylogenetic analyses, we used all publicly available 271 CO1 sequences for individuals identified to species-level. Partial 16S sequences are only available for 10 alvinocaridid species (<u>Table 4</u>). For the present phylogenetic analyses, we used all 29 sequences available in the GeneBank.



Table 1. List of all valid species of the family Alvinocarididae, with remarks on their former and current status.

	Genus		Species	Description, author and	Type locality, depth
Before (Komai, Tsuchida, 2015)	After (Komai, Tsuchida, 2015)	Here		year	
Alvinocari-dinides	Alvinocari-dinides	Alvinocari- dinides	formosa	Komai, Chan, 2010	Gueishandao, Yilan County, Taiwan, 24°51.231'N 0121°59.204'E, 252–275 m
Alvinocaris	Alvinocaris	Alvinocaris	alexander	Ahyong, 2009	Rumble V Seamount, 36°08.27–07.96'S 78°11.74–11.70'E, 485–415 m
Alvinocaris	Alvinocaris	Alvinocaris	brevitel- sonis	Kikuchi, Hashimoto, 2000	"Depression C" of the Minami-Ensei Knoll, 28° 23.35'N 127°38.38'E, 705 m
Alvinocaris	Alvinocaris	Alvinocaris	chelys	Komai, Chan, 2010	Gueishandao, Yilan County, Taiwan, 24°49.682'N 122°0.254'E, 300–276 m
Alvinocaris	Alvinocaris	Alvinocaris	dissimilis	Komai, Segonzac, 2005	Depression C, Minami-Ensei Knoll, 28°23.35'N 127° 38.38'E, 705 m
Alvinocaris	Alvinocaris	Alvinocaris	komaii	Zelnio, Hourdez, 2009	Kilo Moana, Eastern Lau Spreading Center, Lau Basin, southwest Pacific,; 20°9'S 76°12'E, 2620 m
Alvinocaris	Alvinocaris	Alvinocaris	longi-rostris	Kikuchi, Ohta, 1995	Iheya Ridge, Clam Site, Okinawa Trough, 27° 32.70'N 126°58.20'E, 1360 m
Alvinocaris	Alvinocaris	Alvinocaris	lusca	Williams, Chace, 1982	Galapagos Rift Rose Garden area, 0°48.25'N 86° 13.48'W, maximum of 2450 m
Alvinocaris	Alvinocaris	Alvinocaris	markensis	Williams, 1988	Mid-Atlantic Rift Valley about 70 km south of Kane Fracture Zone, 23°22.09'N 44°57.12'W, 3437 m
Alvinocaris	Alvinocaris	Alvinocaris	methano- phila	Komai, Shank, Van Dover, 2005	ODP site 996, Blake Ridge Diapir, 32°29.623'N 76° 11.467'W, 2155 m
Alvinocaris	Alvinocaris	Alvinocaris	muricola	Williams, 1988	West Florida Escarpment, 26°01'N 84°54.61'W, 3277 m
Alvinocaris	Alvinocaris	Alvinocaris	niwa	Webber, 2004	Rumble V, 36°8.63–8.57'S 178°11.77–11.50'E, 877–655 m
Alvinocaris	Alvinocaris	Alvinocaris	stactophila	Williams, 1988	north central Gulf of Mexico about 129 km S of Louisiana, 27°46.94'N 91°30.34'W, 534 m
Alvinocaris	Alvinocaris	Alvinocaris	solitaire	Yahagi, Watanabe, Kojima, Beedessee, Komai, 2014	Central Indian Ridge, Solitaire hydrothermal vent field,19°33.413'S, 65°50.888'E, 2606 m
Alvinocaris	Alvinocaris	Alvinocaris	williamsi	Shank, Martin, 2003	Menez Gwen hydrothermal field, North Atlantic Ocean, 37°50.5'N 31°31.3'W, 850 m
Chorocaris	Chorocaris	Rimicaris	chacei	(Williams, Rona, 1986)	TAG Hydrothermal Field, Mid-Atlantic Ridge, 26° 08.3'N 44°49.6'W, 3620–3650 m
Chorocaris	Chorocaris	Rimicaris	paulexa	Martin, Shank, 2005	Homer Vent (347OC black smoker), 17°37.220'S 113°15.123'W, 2595 m, southern East Pacific Rise
-	Chorocaris	Rimicaris	parva	Komai, Tsuchida, 2015	Manus Basin, South Su, Wave Mercury 2007 (Luk Luk) Campaign, 03°08.09'S, 152°10.5'E, 1310 m
Chorocaris	Chorocaris	Rimicaris	van- doverae	Martin, Hessler, 1990	Alice springs vent field, Mariana Back-Arc Basin, 18°12.599'N 144°42.431'E, 3640 m
-	Chorocaris	Rimicaris	variabilis	Komai, Tsuchida, 2015	Manus Basin, South Su, Wave Mercury 2007 (Luk Luk) Campaign, 03°08.09'S, 152°10.5'E, 1310 m
Mirocaris	Mirocaris	Mirocaris	fortunata	(Martin, Christiansen, 1995)	Vent site 3, Lucky Strike hydrothermal vent, Azores, 37°17.6'N 32°16.5'W, 1624 m
Mirocaris	Mirocaris	Mirocaris	indica	Komai, Martin, Zala, Tsuchida, Hashimoto, 2006	Central Indian Ridge, Kairei Field, 25°19.2'S 70° 02.4'E, 2422 m
-	Manuscaris	Manuscaris	acumi- natus	Komai, Tsuchida, 2015	Manus Basin, South Su, Wave Mercury 2007 Campaign, 03°08.09'S, 152°10.5'E, 1310 m
Nautilocaris	Nautilocaris	Nautilocaris	saint- laurentae	Komai, Segonzac, 2004	North Fiji Basin, White Lady site, 16°59.50'S 173° 55.47'E, 2000 m
Opaepele	Opaepele	Opaepele	loihi	Williams, Dobbs, 1995	Loihi Seamount, Hawaii, 18°55'N 155°16'W, 980 m

(Continued)



Table 1. (Continued)

	Genus		Species	Description, author and	Type locality, depth
Before (Komai, Tsuchida, 2015)	After (Komai, Tsuchida, 2015)	Here		year	
Opaepele	Chorocaris	Rimicaris	susannae	Komai, Giere, Segonzac, 2007	Lilliput, southern Mid-Atlantic Ridge, 09°32.845'S 13°12.546'W, 1500 m, mussel field with diffuse vent fluids
Opaepele	Opaepele	Keldyshi- caris	vavilovi	Lunina, Vereshchaka, 2010	Mid-Atlantic Ridge, Broken Spur vent site, stn 4797
Rimicaris	Rimicaris	Rimicaris	exoculata	Williams, Rona, 1986	TAG Hydrothermal Field, Mid-Atlantic Ridge, 26° 08.3'N 44°49.6'W, 3620–3650 m
Rimicaris	Rimicaris	Rimicaris	hybisae	Nye, Copley, Plouviez, 2011	Mid-Cayman Spreading Centre, Caribbean, Von Damm vent field, 18822.605'N81847.875'W; 2300 m
Rimicaris	Rimicaris	Rimicaris	kairei	Watabe, Hashimoto, 2002	The Central Indian ridge, Indian Ocean, the Kairei Field, 25°19.16'S 70°02.40'E, 2454 m
Shinkaicaris	Shinkaicaris	Shinkaicaris	leurokolos	(Kikuchi, Hashimoto, 2000)	"Depression C" of the Minami- Ensei Knoll, 28° 23.35'N 127°38.38'E, 705 m

Analysis of molecular data

Multiple alignments were made with the use of the Clustal W algorithm [39]. Six CO1 sequences were discarded after alignment, as they represented non-barcoding parts of the CO1 gene or were too short. The remaining 265 aligned sequences were trimmed according to the shortest sequences (Accession Numbers: KC840928-KC840940, HM125910-HM125956) with a total length of 471 bp. Amino acid sequences received from the nucleotide sequences had no stop codons within the open reading frame using the invertebrate mitochondrial code. All 16S sequences after alignment were trimmed according to the shortest sequences (Accession Numbers: AM087916- AM087925) with a total length of 286 bp.

Phylogenetic analysis was performed using both Maximum Likelihood (ML) and Bayesian analyses. To root the resultant trees, *Acanthephyra purpurea* Awas used (also see section 2.2). The best-fit model selected using jModelTest 2.1.7 [40] was the Tamura-Nei model with a gamma distribution and invariable sites (TrN+G+I) for CO1 and the Hasegawa-Kishino-Yano model with a gamma distribution (HKY+G) for 16S data set. These models were used to generate ML gene trees in MEGA 5. Support for branches was assessed using bootstrap analyses with 1,000 replicates [41]. Bayesian phylogenetic analysis was made with the use of MrBayes v3.2.1 [42]. A general time-reversible model (GTR) of sequence evolution with a gamma distribution and invariable sites for CO1 data set was chosen as it represents the closest approximation of the Tamura—Nei model in MrBayes. HKY+G model was used for 16S data set. The Markov Chain Monte Carlo (MCMC) analysis was further used with the following settings: (1) for CO1–18 million generations, trees sampled every 5000 generation, and the first 900 trees discarded; (2) for 16S–1.5 million generations, trees sampled every 1000 generation, and the first 375 trees discarded. The average standard deviation of split frequencies between two runs of MCMC was less than 1% for each analysis, thus indicating convergence.

Estimation of clade robustness

For morphological analyses, we considered the clades robust if they received simultaneous Bremer support ≥ 3 after both analyses. For molecular analyses, we considered the clades robust if they received Bayesian posterior probability value 75%.



Table 2. List of morphological characters and their states.

No	Character	Character state	State No	Figure
	CARAF	ACE		
0	Rostrum	absent	0	1A
		present	1	1B
1	Rostrum	not reaching end of 1st antennular segment	0	1B
		reaching end of 1st antennular segment	1	1C
		overreaching end of 2nd antennular segment	2	1D
2	Rostrum,	tip acute	0	1C
		tip obtuse	1	1B
3	Rostrum	laterally compressed	0	1C
		not laterally compressed	1	1B
4	Rostrum	not dorsoventrally compressed	0	1C
		dorsoventrally compressed	1	1B
5	Rostrum	dorsally carinate	0	1C
		not dorsally carinate	1	1B
6	Rostrum	dorsally smooth	0	1E
		dorsally notched	1	1F
		dorsally toothed	2	1C,D
7	Rostrum, minimal number of dorsal teeth or notches	0	0	
		5–10	1	
		11–15	2	
		16 or more	3	
8	Rostrum, maximal number of dorsal teeth or notches	0	0	
		4–10	1	
		11–15	2	
		16 or more	3	
9	Rostrum	ventrally carinate	0	1C
		not ventrally carinate	1	1B
10	Rostrum	ventrally smooth	0	1E
		ventrally notched	1	1F
		ventrally toothed	2	1D
11	Rostrum, minimal number of ventral teeth or notches	0	0	
		1–2	1	
		3–6	2	
12	Rostrum, maximal number of ventral teeth or notches	0	0	
		1–2	1	
		6–11	2	
13	Rostrum, minimal number of teeth or notches on carapace	0	0	
		1–5	1	
		6–10	2	
14	Rostrum, maximal number of teeth or notches on carapace	0	0	
		1–5	1	
		6–10	2	
15	Carapace, postrostral dorsal carina extending beyond the midlength	absent	0	
		present	1	
16	Carapace, antennal angle	blunt	0	
		acute	1	
17	Carapace, acute pterygostomial tooth	absent	0	

(Continued)



Table 2. (Continued)

No	Character	Character state	State No	Figure
		present	1	
18	Dorsal organ under carapace	absent or inconspicuous	0	
		conspicuous	1	
19	Dorsal organ under carapace	restricted to postorbital region	0	
		extended beyond the postorbital region	1	
20	Dorsal organ	nearly entire	0	
		four-lobed, without pores	1	
		four-lobed, with a pore	2	
	ABDOMEN AN	ID TELSON		
21	Third abdominal segment, posterior margin of pleura	smooth	0	2A
		serrated	1	2B
22	Telson, long linear row of movable dorsolateral spines (≥5 in row)	absent	0	
		present	1	
23	Telson, long sinuous row of movable dorsolateral spines (≥5 in row)	absent	0	
		present	1	
24	Telson, number of strong spines on posterior margin	2–4	0	2C
		6 or more	1	2D
25	Telson, posterior margin	convex	0	2C
		concave	1	2D
26	Telson, posterior concave margin	with shallow incision	0	2D
		nearly bilobed	1	
	THORACIC AP	PENDAGES		
27	Eyestalks	not fused partly	0	
		fused partly, mould seam present	1	
28	Eyestalks	not fused entirely	0	
		fused entirely, without mould seam	1	
29	Eyes, anterior margin	entire	0	
		with conspicuous tubercle	1	1C
30	Antenna II	not operculiform	0	
		operculiform	1	
31	Maxilla II, plumose bacteriophorous setae on scaphognathite	absent	0	2E
		present	1	2F
32	Maxilliped III, epipod	subtriangular	0	
		strap-like	1	ЗА
33	Maxilliped III, epipod	not terminated in hook	0	
		terminated in hook	1	ЗА
34	Maxilliped III, merus	unarmed	0	
		with 1-2 distal spines	1	
35	Pereopod I, epipod	absent or rudimentary	0	
		strap-like	1	ЗА
36	Pereopod I, epipod	not terminated in hook	0	
		terminated in hook	1	ЗА
37	Pereopod I, grooming apparatus	absent or inconspicuous	0	
		conspicuous	1	
38	Pereopod II, epipod	absent or rudimentary	0	
		strap-like	1	ЗА
39	Pereopod II, epipod	not terminated in hook	0	

(Continued)



Table 2. (Continued)

No	Character	Character state	State No	Figure
		terminated in hook	1	ЗА
40	Pereopod II, movable spines on ischium	absent	0	
		present	1	
41	Pereopod III, epipod	absent or rudimentary	0	
		strap-like	1	ЗА
42	Pereopod III, epipod	not terminated in hook	0	
		terminated in hook	1	ЗА
43	Pereopod III, strong movable spines on ischium	absent	0	
		present	1	3B
44	Pereopod III, proximal strong movable spines on merus	absent	0	
		present	1	3B
45	Pereopod III, distal movable spines on merus	absent	0	
		present	1	3B
46	Pereopod III, dactyl	single row of accessory spinules absent	0	
		single row of accessory spinules present	1	
47	Pereopod III, dactyl	two or more rows of accessory spinules absent	0	
		two or more rows of accessory spinules present	1	
48	Pereopod IV, epipod	absent or rudimentary	0	
		strap-like	1	ЗА
49	Pereopod IV, epipod	not terminated in hook	0	
		terminated in hook	1	ЗА
50	Pereopod IV, strong movable spines on ischium	absent	0	
		present	1	зС
51	Pereopod IV, proximal strong movable spines on merus	absent	0	
		present	1	3C
52	Pereopod IV, distal movable spines on merus	absent	0	
		present	1	3C
53	Pereopod IV, dactyl	single row of accessory spinules absent	0	
		single row of accessory spinules present	1	
54	Pereopod IV, dactyl	two or more rows of accessory spinules absent	0	
		two or more rows of accessory spinules present	1	
55	Pereopod V, strong movable spines on ischium	absent	0	
		present	1	3D
56	Pereopod V, dactyl	single row of accessory spinules absent	0	
	•	single row of accessory spinules present	1	
57	Pereopod V, dactyl	two or more rows of accessory spinules absent	0	
	ABDOMINAL API			
58	Pleopod II, appendix interna	developed	0	-
		much reduced	1	3E
59	Pleopod III, appendix interna	developed	0	-
	. , , , ,	much reduced	1	3E
60	Pleopod IV, appendix interna	developed	0	-
	· · · · · · · · · · · · · · · · · · ·	much reduced	1	3E
61	Uropodal exopod, a single movable spine mesial to posterolateral tooth	absent	0	-
	2.2. 2.2. 3.0. post, a olingio monacio opinio modiano postorolatola todin	present	1	-
62	Uropodal exopod, two movable spines mesial to posterolateral tooth	absent	0	_
	participation of the participa	present	1	



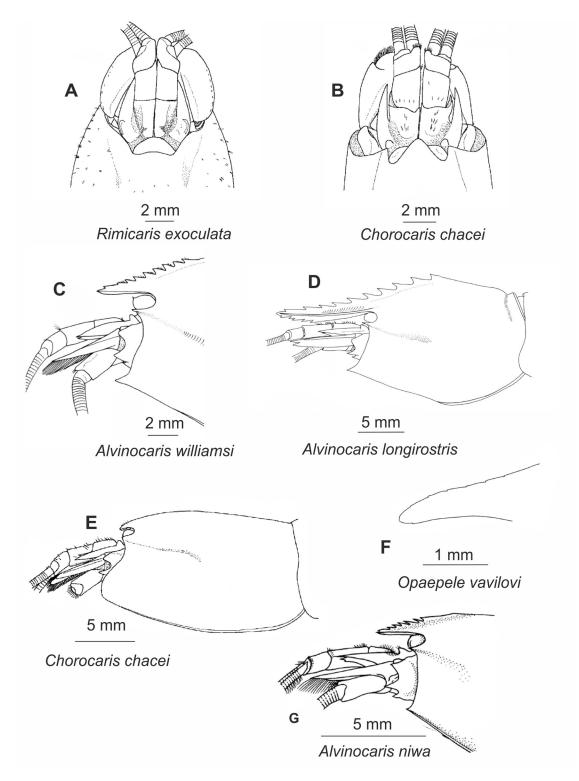


Fig 1. Morphological characters used for phylogenetic analysis. Anterior part of body. (A)-(E) after [13], (F) after [20], (G) after [70].



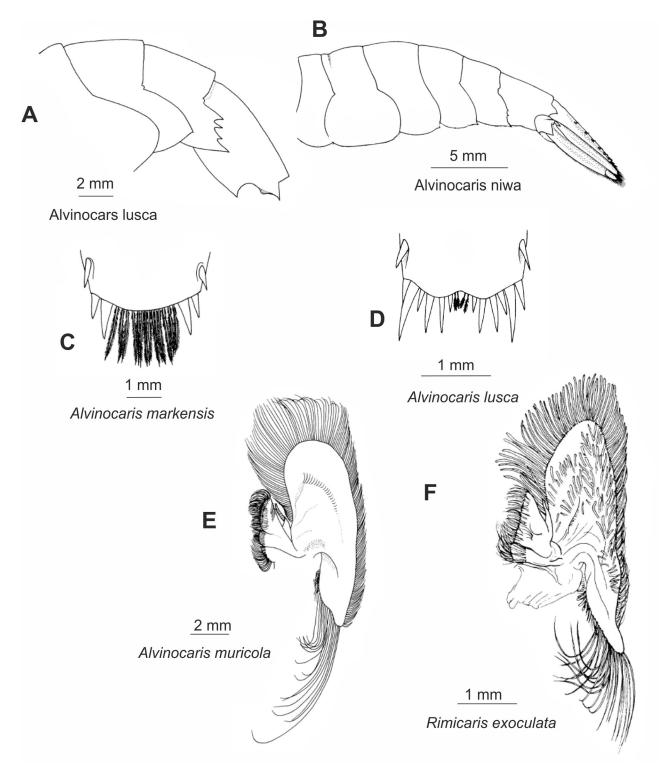


Fig 2. Morphological characters used for phylogenetic analysis. Posterior part of body and maxilla. (A), (C)-(E) after [13], (B) after [70], (F) after [9].



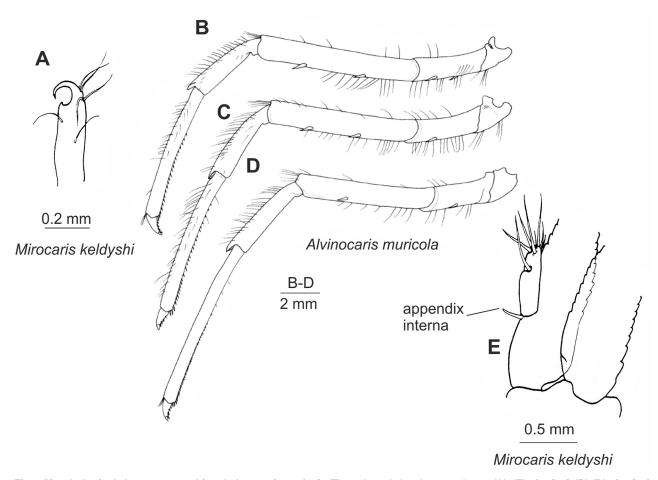


Fig 3. Morphological characters used for phylogenetic analysis. Thoracic and pleonic appendages. (A), (E) after [12], (B)-(D) after [13].

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Results

Morphological clades

Analysis 1 with *Acanthephyra purpurea* as the outgroup retrieved 36 minimal length trees of length 145 (Fig 4A). The basal clade *Alvinocaris* (pink in Fig 4A) forms a sister clade to the rest of the family and is followed by *Keldyshicaris* and two sister clades: *Nautilocaris*+ *Mirocaris* (blue) and *Opaepele+Alvinocaridinides+Manuscaris+Shinkaicaris+Rimicaris* (light green). Within the clade *Alvinocaris*, *Alvinocaris niwa* is the most basal, followed by the other species of *Alvinocaris*. Within the light green clade, there are three clades: *Opaepele*, *Alvinocaridinides*



Table 3. The data matrix of morphological characters of Alvinocarididae.

Species	States of characters
Acanthephyra purpurea	120101 2 111 2 22000100-0000 0-0000010110110110 1 111010 1 111011000000
Alpheus echiurophilus	100111 0 001 0 00000000-0000 0-0010011011011111 1 000000 1 000010000010
Alvinocaris alexander	110000 2 130 2 11121110-0100 0-1010000100100100[01]111000 1 111001000010
Alvinocaris brevitelsonis	120000 2 220 2 12111110-0100 0-101000010010010 1 111000 1 111001000010
Alvinocaris chelis	110000 2 230 2 01121110-0100 0-1010000100100100 0 111000 0 111001000010
Alvinocaris dissimilis	120000 2 230 2 11121110-0100 0-101000010010010 1 111000 1 1110? 1000010
Alvinocaris komaii	120000 2 120 2 12121110-0101 1 1100000010010010 1 110100 1 110110
Alvinocaris longirostris	120000 2 120 2 22121110-0100 0-10100001001001 1 111000 1 111001000010
Alvinocaris lusca	110000 2 120 2 12111110-0101 0-1010000100100100 1 111000 1 111011000010
Alvinocaris markensis	110000 2 230 2 22121110-1101 1 0101000010010010 1 111000 1 1110110
Alvinocaris methanophila	110000 2 230 2 22121110-0100 0-101000010010010 1 111000 1 111001000010
Alvinocaris muricola	120000 2 130 2 22121110-0100 0-101000010010010 1 111000 1 111011000010
Alvinocaris niwa	110000 2 110[02]11001110-1100[01]01010000100100100 1 111000 1 1110? 1000010
Alvinocaris williamsi	110000 2 120 0 00121110-0101 0-101000010010010 1 111000 1 1110? 1000010
Alvinocaris solitaire	120000 2 330 2 11221110-1101 1 0101000010010010 1 111000 1 1110110
Alvinocaris stactophila	110000 2 230 2 11221110-0101 0-101000010010010 1 111000 1 1110? 1000010
Alvinocaridinides formosa	100111 2 231 0 00000111-0010 0-0100000100100000[01]000100[01]00010010010
Rimicaris chacei	101111 0 001 0 00000001100010 0-0100000100100000 0 000100 0 000100100001
Rimicaris paulexa	101111 0 001 0 00000011100010 0-0100000100100000 0 000100 0 000100100001
Rimicaris vandoverae	101111 0 001 0 00000011100010 0-0100000100100000 0 000100 0 000100100001
Rimicaris parva	101111 0 001 0 00000101100010 0-0100000100100000 0 000100 0 000100100001
Rimicaris variabilis	101111 0 001 0 00000111100010 0-0100000100100000 0 000100 0 000100100001
Rimicaris susannae	101111 0 001 0 00000101100010 0-0100000100100000 0 000100[01]000100100001
Rimicaris exoculata	0-1111 0 001 0 00000001110010 0-0101100000000
Rimicaris kairei	0-1111 0 001 0 00000001110010 0-0101100000000
Rimicaris hybisae	101111 0 001 0 00000001120010 0-0100100100100000 0 000100 0 000100100001
Rimicaris loihi	101111[01]011 1 01000111001010 0-0100000100100000 0 000100 0 000100100010
Shinkaicaris leurokolos	110100 2 111 0 00110111000010 0-0100000100100000 0 000100 0 00010010010
Manuscaris acuminata	110100 2 221 0 00110111001100 0-0100000100100100 1 000100 0 00010010010
Mirocaris fortunata	101111 0 001 0 00000111000011 0-1000011111111
Mirocaris indica	101111 0 001 0 00000111000011 0-1000011111111
Nautilocaris saintlaurentae	110111 2 011 0 01000111001011 0-100001111111111
Keldyshicaris vavilovi	101111 1 111 1 01000111001101 0-1010000100100100 1 001000 1 001011000010

Missing data indicated by question marks (?); inapplicable data by hyphens (-); polymorphism indicated by brackets [01]

doi:10.1371/journal.pone.0129975.t003

+*Manuscaris*+*Shinkaicaris*, and *Rimicaris* (green). After discard of all unsupported clades with Bremer support <3, all considered coloured clades persist (Fig 5A).

Analysis 2 with *Alpheus echiurophilus* as the outgroup retrieved 437 minimal length trees of length 146; the tree topology slightly differs from that in Analysis 1, but the principal clades are the same (Fig 4B, same colours). The clade *Nautilocaris*+ *Mirocaris* is basal and followed by three clades: (1) *Opaepele+Alvinocaridinides+Manuscaris+Shinkaicaris+Rimicaris*, (2) *Keldy-shicaris*, and (3) *Alvinocaris*. After discard of all unsupported clades with Bremer support <3, all considered coloured clades persist (Fig 5B).



Table 4. GenBank accession numbers for COI and 16S sequences of species used for phylogenetic analyses.

Species	NCBI GenBank Accession number, Locality	Reference
Alvinocaris dissimilis	AB779491-AB779494	[<u>72</u>],[<u>73</u>]
Alvinocaris komaii	EU031816 Eastern Lau Spreading Center, southwest Pacific	[<u>18</u>]
Alvinocaris longirostris	AB222050, AB222051 Hatoma Knoll, Okinawa Trough	[73], [22]
	NC020313, JQ035659 Hatoma Knoll, Okinawa Trough	[<u>25</u>]
	AB821296 Hatoma Knoll, Okinawa Trough	[72]
	GQ131897	[74]
Alvinocaris lusca	AF125404-AF125407 Galapagos hydrothermal vent field; 9°50'N hydrothermal vent field	[<u>17</u>]
Alvinocaris markensis	KC840879-KC840886, KC840893 Logatchev, Mid-Atlantic Ridge (MAR)	[44]
	AF125408, AF125409 Snake Pit, MAR	[17]
Alvinocaris muricola (including Alvinocaris aff. muricola)	KC840887-KC840892, KC840894-KC840927 Gulf of Mexico, GC852 site; Regab, West Africa	[<u>45</u>]
	EU031814, EU031815 Gulf of Mexico	[75]
	EU868627, EU868628	[<u>76</u>]
Alvinocaris solitaire	LC007114 Solitaire hydrothermal vent field, Central Indian Ridge	[69]
Alvinocaris stactophila	AF125410, AF125411 Louisiana Slope, Gulf of Mexico	[17]
Alvinocaris chelys	NC 018778, JX184903 vent field off Gueishandao (or Kueishan Island), Yilan County, northeastern Taiwan	[77]
Alvinocaris methanophila	AY163260 the Blake Ridge Diapir, Caribbean	[45],[69]
Chorocaris chacei	AF125395-AF125397, AF125414, AF125415 Snake Pit, TAG, MAR	[<u>17</u>]
	KC840928-KC840940 Logatchev, Lucky Strike, MAR	[45]
	AM076957, Lucky Strike, MAR	[78]
	AM087920- AM087922	[79]
Chorocaris parva	AB772278, AB772282 PACMANUS, Manus Basin, South West Pacific	[21]
Chorocaris vandoverae	AF125417, AF125418 Alice Springs, Mariana Back-Arc Basin	[17]
Chorocaris variabilis	AB772279-AB772281, AB7722 PACMANUS, Manus Basin, South West Pacific	[21]
Mirocaris fortunate (including Mirocaris keldyshi)	AF125424-AF125429, AF125430-AF125433 Lucky Strike, Broken Spur, Menez Gwen, TAG, Logatchev, MAR	[17]
	FJ769225, FJ769226 Menez Gwen, MAR	[23]
	AM076959, Lucky Strike, MAR	[<u>78</u>]
	AM087916- AM087919	[79]
Nautilocaris saintlaurentae	NC021971, KF226726 vent the Tofua Arc (Tonga)	[80]
Opaepele loihi	DQ328819-DQ328838 Marianca Arc	[81]
	AF125436, AF125437 Loihi Seamount	[17]
	NC020311, JQ035657 Nikko Seamount, Philippine Sea Plate	[25]
Rimicaris exoculata	AF125398-AF125403, AF125419, AF125420, AF125440 TAG, Lucky Strike, Broken Spur, Rainbow	[17]
	AF044057 MAR	[82]
	FN392996-FN393005 Rainbow, TAG, Logatchev, South MAR	[24]
	HM125910-HM125956 Rainbow, TAG, Logatchev, Ashadze, South MAR	[83]
	AF035459, TAG, South MAR	[82]
	AM087923-AM087925	[79]
	AM076958, Lucky Strike, MAR	[78]
Rimicaris kairei	AB813087-AB813108 Dodo, Solitaire, Edmond and Kairei fields	[84]
	NC020310, JQ035656 Kairei Field (Rodriguez Triple Junction)	[25]
	JN850606, JN850607 Beebe and Von Damm vent fields (Caribbean)	[3]
Rimicaris hvbisae		L=1
Rimicaris hybisae		[85]
Rimicaris hybisae Acanthephyra purpurea	KJ566968-KJ5678003 Beebe and Von Damm vent fields (Caribbean) GU183787, GU183788	[<u>85</u>]



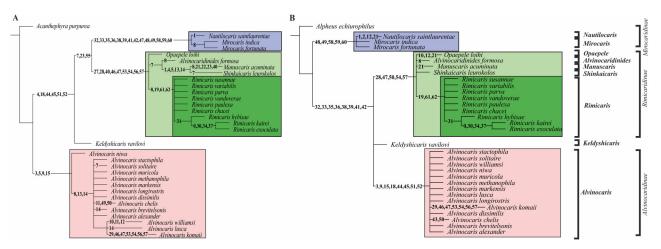


Fig 4. Strict consensus trees of Alvinocarididae and synapomorphies (numbers in circles). A, analysis 1 with *Acanthephyra purpurea* as the outgroup. B, analysis 2 with *Alpheus echiurophilus* as the outgroup. Supported clades are marked by different colors: Mirocaridinae (blue), Alvinocaridinae (pink), Rimicaridinae (light green), *Rimicaris* (green).

Molecular clades

The molecular phylogenetic Analysis 1 with use of CO1 gene resulted in a tree (Fig 6A), which resembles the morphological tree except the branching of *Alvinocaris* (Fig 5B). In the morpho analysis *Alvinocaris* forms a single clade (not internally resolved), but in the genetic analysis three (COI) or two (16S) clades occur.

The clade Mirocaris+*Nautilocaris* (Fig 6A, coloured in blue) is supported both by ML bootstrap (98) and Bayesian posterior probability (100). This clade comprises two genera: *Mirocaris* (*M. fortunata*) and *Nautilocaris* (*N. saintlaurentae*). Genetic difference between the genera

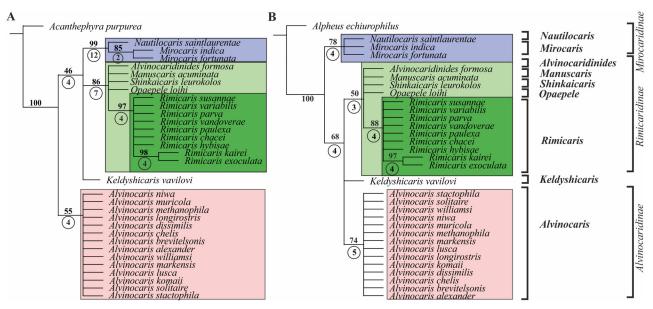


Fig 5. Statistically robust clades of Alvinocarididae with bootstrap support (numbers above the clade) and Bremer support (numbers below the clade in circles). A, analysis 1 with *Acanthephyra purpurea* as outgroup. B, analysis 2 with *Alpheus echiurophilus* as outgroup. Supported clades are marked by different colors: Mirocaridinae (blue), Alvinocaridinae (pink), Rimicaridinae (light green), *Rimicaris* (green).

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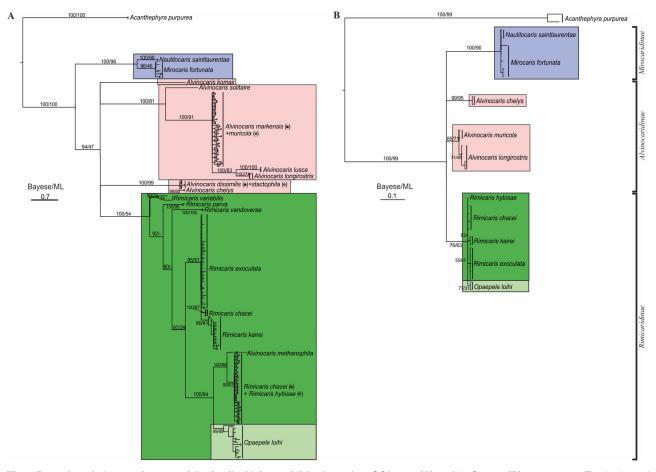


Fig 6. Bayesian phylogenetic trees of the family Alvinocarididae based on COI gene (A) and 16 S gene (B) sequences. The horizontal scale bar marks the number of expected substitutions per site. Statistical support indicated as Bayesian posterior probabilities (left values) and bootstrap analysis with 1,000 replicates (right values) and. Supported clades are marked by different colors: Mirocaridinae (blue), Alvinocaridinae (pink), Rimicaridinae (light green), Rimicaris (green).

reaches 5.1%. This clade is identical to the clade *Nautilocaris*+ *Mirocaris* revealed by both morphological analyses. The basal clade is followed by four clades.

The clade *Alvinocaris komaii* (Fig 6A, pink) comprises only one species of *Alvinocaris*. Position of this on the tree topology is unresolved and thus requires further research.

The clade Alvinocaris solitaire+Alvinocaris markensis+Alvinocaris muricola+ Alvinocaris lusca+Alvinocaris longirostris (Fig 6A, pink) is supported by ML bootstrap (81) and Bayesian posterior probability (100). This clade comprises a significant part of the genus Alvinocaris (Fig 6A, coloured in pink). Alvinocaris solitaire is basal and followed by A. markensis, A. muricola and Alvinocaris lusca+Alvinocaris longirostris. Genetic differences between three terminal subclades range from 5 to 6.7%.

The clade Alvinocaris dissimilis +Alvinocaris stactophila +Alvinocaris chelis (Fig 6A, pink) is supported by ML bootstrap (99) and Bayesian posterior probability (100). This clade comprises the rest of Alvinocaris used in Analysis 1 (Fig 6A, coloured in pink). The former two species are genetically identical, while the genetic distance (p-distance) between them and A. chelys reaches 0.5%.

The clade Rimicaris variabilis + Rimicaris parva + Rimicaris vandoverae + Rimicaris exoculata + Rimicaris chacei + Rimicaris kairei + Alvinocaris methanophila + Rimicaris



hybisae+ Opaepele loihi (Fig 6A, light green) is supported by Bayesian posterior probability (100). This clade comprises a part of the genus *Rimicaris* (Fig 6A, coloured in green) and two other genera: *Alvinocaris* (a single sequence) and *Opaepele*. Specimens of *C. chacei* are divided into two distant groups with genetic differences 7.9%.

The molecular phylogenetic Analysis 2 with use of 16S gene resulted in a tree with similar clades (Fig 6B). The monophyly of the clades *Mirocaris+Nautilocaris* and *Rimicaris exoculata + Rimicaris chacei + Rimicaris kairei + Rimicaris hybisae+* was supported (Bayesian posterior probability (100 and 76, respectively). *Alvinocaris* created two clades *Alvinocaris longiros-tris+Alvinocaris muricola* and *Alvinocaris chelis* (Bayesian posterior probability 85 and 99, respectively), which agree with two clades of Analysis 1

Discussion

Both morphological analyses revealed three major robust monophyletic clades, herein assign to them subfamiliar status, as Mirocaridinae, Rimicaridinae subfam. nov. and Alvinocaridinae subfam. nov.

The clades

The clade/subfamily *Mirocaridinae* (Fig 5A and 5B, blue) comprises three species and two genera: *Nautilocaris* and *Mirocaris*. This clade is supported by the following synapomorphies common for both morpho analyses (starting from the character number):

(48-49) presence of strap-like epipods terminating in a hook, on the fourth pereopod,

(58-60) much reduced appendix interna in second to fourth pleopods

The clade is well supported by molecular data (Fig 6A).

The clade/subfamily *Rimicaridinae* (Fig <u>5A</u> and <u>5B</u>, light green) comprises thirteen species and five genera: *Opaepele*, *Alvinocaridinides*, *Manuscaris*, *Shinkaicaris*, and *Rimicaris*. This clade is also supported by the following synapomorphies common for both morpho analyses:

(28) entirely fused eyestalks without mould seam,

(47, 54, 57) presence of two or more rows of accessory spinules on the dactyls of the third to fifth pereopods.

The clade is supported by molecular data (Fig 6A). The position of *A. methanophila* within Rimicaridinae is worthy of comment. This result is based on a single specimen of *A. methanophila*, which was collected at the Blake Ridge Diapir site, sequenced and deposited in GenBank (Accession No AY163260) under the name «*Alvinocaris* sp. TMS-2002». The shrimp was originally named «Blake Ridge shrimp» [43]. Later this specimen along with 33 other adult specimens was described as a new species *A. methanophila* [44]. Texeira et al [45] used the same sequence and included this in their clade "ESU 2" which comprised specimens from three genera. We feel that the resulting position of *A. methanophila* in a common clade with *R. chacei* and *R. hybisae* is a result of incorrect identification or processing the material.

Molecular analyses indicate close relation between *Opaepele (O. loihi)* and *Rimicaris*, however morphologically they are quite distinct and for now we keep both genera as valid. Both morphological analyses supported validity of remaining genera of the clade Rimicaridinae: *Alvinocaridinides*, *Shinkaicaris*, and *Manuscaris*.

The clade *Rimicaris* is nested within the clade Rimicaridinae and comprises all species of the currently recognised genera *Chorocaris* and *Rimicaris* (Fig <u>5A</u> and <u>5B</u>, green). This clade received very high support during morphological and molecular analyses and is also supported by the synapomorphies common for both morpho analyses:

- (19) dorsal organ under carapace extended beyond the postorbital region,
- (61-62) presence of two movable spines mesial to posterolateral tooth on uropodal exopod.



As this is a robust monophyletic clade (except terminal *R. exoculata+R. kairei*), we herein synonymize *Chorocaris* with *Rimicaris*, with Rimicaris being the older name. We note that the type species of *Chorocaris*, *Chorocaris chacei*, was initially described as *Rimicaris* [9]. Our analyses also show that the former *Opaepele susannae* was correctly removed from the genus *Opaepele* (and transferred to *Chorocaris*) [21].

Recent molecular studies by Texeira et al [45] based on studies from the Tropical Atlantic have previously showed a common clade for *Rimicaris* and *Chorocaris*. "These showed very low genetic divergence at levels similar to divergence between individuals of the same species. We posit that these taxa belong to the same genus, possibly even the same species" [45].

The clade is well supported by our molecular data (Fig 6A). A chequerwise arrangement of the species belonging to the former *Rimicaris* and *Chorocaris* within Rimicaridinae gives additional evidence for their genetic similarity and thus synonimising both genera.

One of the unexpected results of our molecular analyses is the position of specimens of *R. chacei* in two different and well supported clades (distance 7.9%). Those specimens which are closer to *R. exoculata*, were identified and sequenced by T.M. Shank (NN AF125395-397, AF125414-415 from Snake Pit and TAG [17]. The specimens closer to *Opaepele loihi* (NN KC840928-KC840940 from Logatchev and Lucky Strike) are from Teixeira et al. [45]. This division of *R. chacei* into two groups may be caused by various factors:

- mitochondrial introgression in which mitochondrial DNA of *R. exoculata* may have been incorporated in the populations of *C. chacei* at Snake Pit and TAG. Similar effects were found in other marine Malacostraca [46], [47] and also in vent Bivalvia [48], [49],
- existence of cryptic species of *C. chacei* recognized as a single morphological species.

The clade/subfamily *Alvinocaridinae* (Fig <u>5A</u> and <u>5B</u>, pink) comprises fourteen species of the genus *Alvinocaris*. This clade is also supported by the following synapomorphies common for both morpho analyses:

- (3, 9) laterally compressed and ventrally carinate rostrum,
- (15) presence of postrostral dorsal carina extending beyond the midlength of carapace.

Molecular analyses indicate presence of three supported clades of *Alvinocaris* (Fig 6A). Here we consider the genus and the subfamily as monophyletic on the basis of the two morphological analyses and leave the possibility of a polyphyletic origin of the clade to be resolved by more data in the future.

The clade *Keldyshicaris* comprises a single species, *Keldyshicaris vavilovi*. Both morphological analyses did not support monophyly of the former genus *Opaepele* and we suggest division of this genus into two monotypic genera, *Opaepele* with *O. loihi* (subfamily Rimicaridinae) and *Keldyshicaris* gen.n. with *K. vavilovi* n. comb. The status of this genus may be further clarified after receiving molecular data.

Morphological trends in Alvinocarididae

Mirocaridinae and Rimicaridinae share a degenerate rostrum, reduced external spines and the presence of the dorsal organ. This type of rostrum may be advantageous in the vicinity of shimmering waters and vent fluids where Mirocaridinae and Rimicaridinae occur. Predators are rare in these extreme biotopes where sulphides, heavy metals, and methane are actively leaching from surrounding rocks [50]. A shorter unarmed rostrum along with reduced spines reduces impact of strong turbulent water fluxes, which are common in the microbiotopes where the shrimps thrive [51].

The dorsal organ has been described inside the carapaces of all recent genera of Mirocaridinae and Rimicaridinae [2], [20], [52-53]. These spot-like organs are believed to be homologous



to the 'dorsal eye' found in *R. exoculata* [2], [54-55], but are smaller and do not comprise four lobes. The dorsal organ, also called 'dorsal eye', is an extremely efficient photoreceptor, used for detecting light emitted from the vents [56-58].

Mirocaridinae further differ from other Alvinocarididae in having strap-like epipods terminating in a hook and in much reduced appendix interna in second to fourth pleopods. The strap-like pereopodal epipods are common for many caridean families and these characters may be retained from the ancestor. Modification in pleopodal characters may be related to specific traits in movement or copulation, which may be adaptive in the shimmering waters where Mirocaridinae thrive.

Rimicaridinae possess entirely fused eyestalks and presence of two or more rows of accessory spinules on the dactyls of the third to fifth pereopods—adaptations favouring anchoring close to the strong currents hydrothermal fluids. Within the clade Rimicaridinae, the genus *Rimicaris* shows further modification of external structures on the rostrum and frontal part of carapace, further development of the dorsal organ, and elaboration of uropodal exopod. The polarization of the characters 19 along with inflation of carapace and extension of scaphognathite shows deeper association of *Rimicaris* with vent fluids than any of other genera of the subfamily [51],[59], [60]. Presence of two strong movable spines instead of one (the polarization of characters 61–62) may indicate importance of the tail fan, which is used in the escape behaviour of shrimps ([61]. The presence of additional spines may make this behaviour more efficient when high-temperature turbulent water fluxes can suddenly erupt from the rocks and damage shrimps [51].

Alvinocaridinae are characterized by a well-developed instead of reduced rostrum and post-rostral carina, by a frontal ocular tubercle, and by the spination of the fourth-fifth pereopods. These traits are plausibly accounted for by their habitat at the periphery of hydrothermal vent fields [60]. In these habitats, predatory fish do occur and frontal armature may partly protect the shrimps from attacks. The frontal ocular tubercle indicates that the eyes may have additional chemo- or mechanosensory function facilitating orientation at the vent field by means of the frontal tubercle. If so, the tubercle may represent a sensory mechanism alternative to the photoreceptory dorsal organ of other Alvinocarididae. Distal movable spines on the merus of third and fourth pereopods (characters 45 and 51) are present in Alvinocaridinae, potentially enabling more efficient prey catching and sorting and processing the organic particles which the species live on.

Classification of Alvinocarididae

Subfamily Mirocaridinae, subfam. nov

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Diagnosis. Carapace dorsally smooth, without postrostral carina; dorsal organ conspicuous; telson bearing 12–19 strong spines. Eyes partly fused, anterior margin of cornea without developed tubercle; epipods of first to fourth pereopods strap-like, terminating in a hook; meri of third and fourth pereopods without movable spines; appendix interna in pleopods II-V much reduced.

Type genus. Mirocaris Vereshchaka, 1997 [12], by original designation.

Genera included. *Mirocaris* Vereshchaka, 1997 [12], *Nautilocaris* Komai, Segonzac, 2004 [14].

Remarks. The type species of the genus *Mirocaris* is *M. keldyshi*, a junior subjective synonym of *Chorocaris fortunata*. Although the generic status of *Mirocaris* has largely been supported, the family Mirocarididae Vereshchaka, 1997 has not been recognized, but is herein



resurrected at subfamily level [19]. Molecular data have been previously indicating high status of *Mirocaris* [17].

Genus Mirocaris Vereshchaka, 1997 [12]

Diagnosis. Rostrum dorsoventrally compressed, not reaching end of first antennular segment, apically obtuse, dorsally and ventrally not carinate, smooth; carapace with antennal angle acute, pterygostomial tooth present; dorsal organ restricted to postorbital region; third abdominal segment not serrated; telson with sinuous row of movable dorsolateral spines. Anterior margin of cornea without tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with distal movable spine on antepenultimate segment; first pereopod with grooming apparatus; second pereopod with a distal movable spine on ischium; ischia of third to fifth pereopods with or without movable spines; dactyli of third to fifth pereopods with a single row of accessory spinules; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. *Mirocaris keldyshi Vereshchaka*, 1997 (junior subjective synonym of Chorocaris fortunata Martin & Christiansen, 1995) [62].

Species included. *Mirocaris fortunata* (Martin, Christiansen, 1995) [62], *Mirocaris indica* Komai, Martin, Zala, Tsuchida, Hashimoto, 2006 [63].

Remarks. The genus includes 2 species, one from the Atlantic, and the other from the Indian Ocean. Both species are so similar in morphology that supporting molecular data are necessary to prove the validity of M. indica. We keep here, however, a conservative approach and recognize both species.

Genus Nautilocaris Komai, Segonzac, 2004[14]

Diagnosis. Rostrum dorsoventrally compressed, overreaching end of first antennular segment, apically acute, dorsally not carinate, toothed, ventrally smooth; carapace with antennal angle acute, pterygostomial tooth present; dorsal organ restricted to postorbital region; third abdominal segment serrated; telson with sinuous row of movable dorsolateral spines. Anterior margin of cornea without tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with a distal movable spine on antepenultimate segment; first pereopod with grooming apparatus; second pereopod with distal movable spine on ischium; ischia of third to fifth pereopods with or without movable spines; dactyli of third to fifth pereopods with a single row of accessory spinules; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. *Nautilocaris saintlaurentae* Komai & Segonzac, 2004 [14], by monotypy. **Species included.** *Nautilocaris saintlaurentae* Komai & Segonzac, 2004 [14].

Remarks. *Nautilocaris* differs from the closely related genus *Mirocaris* in having a longer and denticulate rostrum and in the serrated pleura of the third abdominal somite.

Subfamily Rimicaridinae, subfam.nov

urn:lsid:zoobank.org:act:1E84ACE4-B031-43BA-8B91-CD0AFB4DBF77

Diagnosis. Carapace dorsally smooth, without postrostral carina; dorsal organ conspicuous; telson bearing 12–19 strong spines. Eyes fused entirely, anterior margin of cornea without developed tubercle; epipods of first to fourth pereopods rudimentary; meri of third and fourth pereopods without movable spines; appendix interna in pleopods II-V developed.

Type genus. *Rimicaris* Williams & Rona, 1986 [9], by present designation.



Genera included. Alvinocaridinides Komai & Chan, 2010 [2], Manuscaris Komai & Tsuchida, 2015 [21], Opaepele Williams & Dobbs, 1995 [11], Rimicaris Williams & Rona, 1986 [9], and Shinkaicaris Komai & Segonzac, 2005 [13].

Genus Alvinocaridinides Komai & Chan, 2010 [2]

Diagnosis. Rostrum dorsoventrally compressed, not reaching end of first antennular segment, apically blunt, dorsally toothed, ventrally smooth; carapace dorsally smooth, antennal angle acute, pterygostomial tooth present; dorsal organ restricted to postorbital region; third abdominal segment not serrated; telson with sinuous row of movable dorsolateral spines, posterior margin bearing 4 strong spines. Anterior margin of cornea without tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with 1 distal movable spine on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with grooming apparatus; second pereopod without distal movable spine on ischium; ischia of third to fifth pereopods with or without movable spines; dactyli of third to fifth pereopods with two or more rows of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. Alvinocaridinides formosa Komai & Chan, 2010 [$\underline{2}$], by original designation and monotypy.

Species included. Alvinocaridinides formosa Komai & Chan, 2010.

Remarks. Komai and Chan [2] established this genus for a single new species, *A. formosa*. Our analyses support the validity of this genus.

Manuscaris Komai & Tsuchida, 2015 [21]

Diagnosis. Rostrum laterally compressed, reaching end of first antennular segment, apically acute, dorsally carinate, toothed, ventrally smooth; carapace dorsally toothed, pterygostomial tooth present; third abdominal segment serrated; telson with linear row of movable dorsolateral spines, posterior margin bearing 2–4 strong spines. Anterior margin of cornea without tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with a distal movable spine on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with grooming apparatus; second pereopod with distal movable spine on ischium; ischia of third to fifth pereopods with or without movable spines; dactyli of third to fifth pereopods with two or more rows of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. *Manuscaris acuminata* Komai and Tsuchida, 2015, by monotypy. **Species included.** *Manuscaris acuminata* Komai and Tsuchida, 2015 [21]. **Remarks.** This genus was recently erected [21] and our analyses support its validity.

Opaepele Williams and Dobbs, 1995 [11]

Diagnosis. Rostrum dorsoventrally compressed, not reaching end of first antennular segment, apically blunt, dorsally and ventrally not carinate, notched or smooth; carapace dorsally smooth, antennal angle acute, pterygostomial tooth present; dorsal organ restricted to postorbital region; third abdominal segment serrated; telson with sinuous row of movable dorsolateral spines, posterior margin bearing 2–4 strong spines. Anterior margin of cornea without tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with a distal movable spine on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with grooming apparatus; second pereopod



without distal movable spine on ischium; ischia of third to fifth pereopods without movable spines; dactyli of third to fifth pereopods with two or more rows of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. *Opaepele loihi* Williams, Dobbs, by monotypy. **Species included.** *Opaepele loihi* Williams, Dobbs, 1995 [11]

Remarks. Opaepele is herein restricted to the type species only, O. loihi. Opaepele susannae has been recently transferred to Chorocaris [21] and our analyses confirm this decision. Opaepele vavilovi is transferred to a new genus, Keldyshicaris (see below).

Genus Rimicaris Williams & Rona, 1986 [9]

Diagnosis. Rostrum if present dorsoventrally compressed, not reaching end of first antennular segment, apically obtuse, dorsally and ventrally not carinate, smooth; carapace dorsally smooth, antennal angle blunt or acute, pterygostomial tooth present or absent; dorsal organ extending beyond the postorbital region; third abdominal segment smooth or serrated; telson with sinuous row of movable dorsolateral spines, posterior margin bearing 2–4 strong spines. Eyes fused entirely, anterior margin of cornea without developed tubercle; scaphognathite expanded, with or without heavily plumose bacteriophore setae; third maxilliped with 0–2 distal movable spines on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with or without grooming apparatus; second pereopod without distal movable spine on ischium; ischia of third to fifth pereopods with or without movable spines; dactyli of third to fifth pereopods with two or more rows of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with two movable spines mesial to posterolateral tooth.

Type species. Rimicaris exoculata Williams & Rona, 1986 [9], by original designation. Species included. Rimicaris chacei (Williams, Rona, 1986) [9], Rimicaris exoculata Williams, Rona, 1986, Rimicaris hybisae Nye, Copley, Plouviez, 2012 [3], Rimicaris kairei Watabe, Hashimoto, 2002 [64], Rimicaris parva (Komai, Tsuchida) [21], Rimicaris paulexa (Martin, Shank, 2005) [65], Rimicaris susannae (Komai, Giere, Segonzac, 2007) [66], Rimicaris vandoverae (Martin, Hessler, 1990) [10], and Rimicaris variabilis (Komai, Tsuchida) [21].

Remarks. The genus includes all species of the former genera *Chorocaris* sensu Komai and Tsuchida [21] and *Rimicaris*.

Rimicaris exoculata from the Atlantic and *R. kairei* from the Indian Ocean are very similar in morphology and are not statistically distinct on the molecular tree (Fig 6A). We keep a conservative approach and recognize both species, while pointing out a need of additional research to confirm their validity.

Rimicaris vandoverae and R. paulexa are so similar morphologically that it is impossible to articulate sharp distinctions between them. The species are geographically isolated and the minor morphological distinctions may refer to an inter-population difference rather than to an inter-specific variability. Molecular data (now missing for R. paulexa) will help in understanding the status of these species.

As stated above, *R. chacei* may include at least two cryptic species. New sequences are desirable to clarify this situation.

Genus Shinkaicaris Komai & Segonzac, 2005 [13]

Diagnosis. Rostrum laterally compressed, overreaching end of first antennular segment, apically acute, dorsally carinate, toothed, ventrally smooth; carapace dorsally toothed, antennal angle acute, pterygostomial tooth present; dorsal organ restricted to postorbital region; third



abdominal segment not serrated; telson with sinuous row of movable dorsolateral spines, posterior margin bearing 2–4 strong spines. Anterior margin of cornea without tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with a distal movable spine on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with grooming apparatus; second pereopod without distal movable spine on ischium; ischia of third to fifth pereopods without movable spines; dactyli of third to fifth pereopods with two or more rows of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. Alvinocaris leurokolos Kikuchi & Hashimoto, 2000 [67], by monotypy. **Genera included.** Shinkaicaris leurokolos Kikuchi, Hashimoto, 2000. **Remarks.** Our analyses support the validity of the genus.

Subfamily Alvinocaridinae, subfam. nov

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Diagnosis. Rostrum laterally compressed, overreaching end of first antennular segment, apically acute, dorsally carinate and toothed, ventrally carinate, toothed or smooth; carapace dorsally toothed or smooth, with postrostral carina extending beyond midlength of carapace, antennal angle acute, pterygostomial tooth present; dorsal organ inconspicuous; third abdominal segment smooth or serrated; telson with linear row of movable dorsolateral spines, posterior margin convex or concave, bearing 4–18 strong spines. Eye partly fused, anterior margin of cornea with developed tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with 1–2 distal movable spines on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with grooming apparatus; second pereopod with distal movable spines on ischium; meri of third and fourth pereopods with movable spines; ischia of third to fifth pereopods with movable spines; dactyli of third to fifth pereopods with a single row of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type genus. Alvinocaris Williams, Chace, 1982 [8], by present designation **Genera included.** Alvinocaris Williams, Chace, 1982 [8].

Genus Alvinocaris Williams, Chace, 1982 [8]

Diagnosis. As in subfamily.

Type species. *Alvinocaris lusca* Williams, Chace, 1982 [8], by monotypy.

Species included. Alvinocaris alexander Ahyong, 2009 [38], A. brevitelsonis Kikuchi, Hashimoto, 2000 [67], A. chelys Komai, Chan, 2010 [2], A. dissimilis Komai, Segonzac, 2005 [13], A. komaii Zelnio, Hourdez, 2009 [18], A. longirostris Kikuchi, Ohta, 1995 [7], A. lusca Williams, Chace, 1982 [8], A. markensis Williams, 1988 [68], A. methanophila Komai, Shank, Van Dover, 2005 [69], A. muricola Williams, 1988 [68], A. niwa Webber, 2004 [70], A. solitaire Yahagi, Watanabe, Kojima, Beedessee, Komai, 2014 [71], A. stactophila Williams, 1988 [68], A. williamsi Shank, Martin, 2003 [70].

Remarks. Molecular data indicate a presence of three species groups at least: (1) *A. komaii*, (2) *A. solitaire*, *A. markensis*, *A. muricola*, *A. lusca*, *A. longirostris*, and (3) A. *dissimilis*, *A. stactophila*, *Alvinocaris chelis*. Morphological analyses do not provide robust clades within *Alvinocaris*.

Keldyshicaris gen.nov

urn:lsid:zoobank.org:act:4D7D81BC-2C9A-45A0-93EC-D276E33FA174



Diagnosis. Rostrum dorsoventrally compressed, not reaching end of first antennular segment, apically blunt, dorsally not carinate, dorsally and ventrally notched; carapace dorsally smooth, antennal angle acute, pterygostomial tooth present; dorsal organ restricted to postorbital region; third abdominal segment serrated; telson with linear row of movable dorsolateral spines, posterior margin bearing 2–4 strong spines. Anterior margin of cornea with rudimentary tubercle; scaphognathite not much expanded, without heavily plumose bacteriophore setae; third maxilliped with a distal movable spine on antepenultimate segment; epipods of first to fourth pereopods rudimentary; first pereopod with grooming apparatus; second pereopod with distal movable spine on ischium; dactyli of third to fifth pereopods with two or more rows of accessory spinules; appendix interna in pleopods II-V developed; uropodal exopod with a single movable spine mesial to posterolateral tooth.

Type species. Opaepele vavilovi Lunina and Vereshchaka, 2010.

Species included. *Keldyshicaris vavilovi* (Lunina and Vereshchaka, 2010) [20].

Etymology. Named after the Russian R/V "Akademik Mstislav Keldysh" which significantly contributed to the studies of vent fauna.

Remarks. The proper position and status of *Keldyshicaris* within Alvinocarididae remains uncertain.

Key to subfamilies, genera, and species of Alvinocarididae (Table 5)

Table 5.

Carapace with conspicuous postrostral carina extending beyond the midlength; no conspicuous dorsal organ. Meri of pereopods III-IV with strong movable spines	2 (subfamily Alvinocaridinae, <i>Alvinocaris</i>
- Carapace without conspicuous postrostral carina extending beyond the midlength; dorsal organ conspicuous; Meri of pereopods III-IV without strong movable spines	10
2. Dorsal teeth present only on rostrum	Alvinocaris niwa
- Dorsal teeth both on rostrum and on carapace	
3. Telson with 2–4 strong spines on posterior margin	4
- Telson with 6 or more strong spines on posterior margin	8
4. No strong movable spines on ischium of fourth pereopods	Alvinocaris chelys
- Ischium of fourth pereopod with strong movable spines	
5. Rostrum not reaching end of second antennular segment	A. methanophyla in the Atlantic and A. alexander in the Pacific Ocean
- Rostrum overreaching end of second antennular segment	6
6. Ventral margin of rostrum with 1–2 small subdistal teeth	Alvinocaris dissimilis
- Ventral margin of rostrum with 3 or more teeth	
7. Posterior margin of telson with >2 pairs of spines	Alvinocaris brevitelsonia
- Posterior margin of telson with 2 pairs of spines at lateral corners and 12–14 plumose setae	A. muricola in the Atlantic and A. longirostris in the Pacific
8. Posterior margin of telson concave	9
- Posterior margin of telson convex	1:
9. Pleura of third abdominal somite smooth; posterior margin of telson bilobed; dactyli of third to fifth pereopods with two or more rows of accessory spinules	Alvinocaris koma
- Pleura of third abdominal somite serated; posterior margin of telson slightly concave; dactyli of third to fifth pereopods with a single row of accessory spinules	10
10. Rostrum not reaching end of second antennular segment, with 3 or more ventral teeth	Alvinocaris markensi
- Rostrum overreaching end of second antennular segment, with 1–2 ventral teeth	Alvinocaris solitaire
11. Rostrum with ventral teeth	12



Table 5. (Continued)

Alvinocaris williams	o ventral teeth on rostrum
Alvinocaris stactophyl	Rostrum with a single ventral tooth. Carapace with 6–10 dorsal teeth
Alvinocaris lusc	ostrum with 2-6 ventral teeth. Carapace with 1-5 dorsal teeth
14 (subfamily Mirocaridinae	Epipods of first to fourth pereopods developed, strap-like, ending in hook; pendices internae of second to fourth pleopods rudimentary
15 (subfamily Rimicaridinae	pipods of first to fourth pereopods rudimentary, not strap-like, not ending in hook; pendices internae of second to fourth pleopods normally developed
Mirocaris (M. fortunata in the Atlantic, M. indica in the India Ocean	Rostrum not reaching end of first antennular segment, dorsally smooth. Pleura of d abdominal somite smooth
Nautilocaris, the only species N. saintlaurenta	ostrum overreaching end of first antennular segment, with dorsal teeth. Pleura of d abdominal somite serrate
1	Rostrum overreaching end of 1st antennular segment, carapace with dorsal teeth
1	ostrum not reaching end of 1st antennular segment, carapace without dorsal teeth
genus Manuscaris, the only species Manuscaris acuminate	Rostrum and carapace bearing a total of 11 or more dorsal teeth; pleura of third dominal somite serrate; row of dorsolateral spines on telson linear; ischia of third and orth pereopods with movable spines
Shinkaicaris, the only species Shinkaicaris leurokolo	strum and carapace bearing a total of 10 or less dorsal teeth; pleura of third lominal somite not serrate; row of dorsolateral spines on telson sinuous; ischia of d and fourth pereopods without movable spines
1:	Rostrum with dorsal teeth or notches; dorsal organ restricted to postorbital region; podal exopod with a single movable spine mesial to posterolateral tooth.
20 (genus <i>Rimicari</i> s	ostrum without dorsal teeth or notches; dorsal organ extending beyond postorbital ion; uropodal exopod with two movable spines mesial to posterolateral tooth
Alvinocaridinides, the only species Alvinocaridinides formosa	Rostrum with acute tip, bearing >10 dorsal teeth, ventrally unarmed; pleura of third dominal somite not serrate
1:	ostrum with blunt tip, bearing <10 dorsal notches, ventrally armed with 1–2 notches; ura of third abdominal somite serrate
Opaepele, the only species Opaepele loih	Telson with sinuous row of dorsolateral spines and 2–4 spines on posterior margin; nea without anterior tubercle; ischia of third and fifth pereopods without strong vable spines; dactyli of third to fifth pereopods with two or more rows of accessory nules
Keldyshicaris, the only species Keldyshicaris vavilou	elson with linear row of dorsolateral spines and >10 spines on posterior margin; nea with anterior tubercle; ischia of third and fifth pereopods with strong movable nes; dactyli of third to fifth pereopods with a single row of accessory spinules
2	Carapace width not exceeding carapace height in adults; dorsal organ nearly ire; scaphognathite without heavily plumose bacteriophore setae
2	arapace width exceeding carapace height in adults; dorsal organ four-lobed; phognathite with heavily plumose bacteriophore setae
2	Dactyli of third to fifth pereopods with two rows of accessory spinules
2	actyli of third to fifth pereopods with 3–4 rows of accessory spinules
Chorocaris parv	Pleura of fourth and fifth abdominal somites not serrated
Chorocaris susanna	eura of fourth and fifth abdominal somites serrated
Chorocaris chace	Pterigostomial tooth absent
24	erigostomial tooth present
Chorocaris variabili	Pleura of fourth and fifth abdominal somites serrated
Chorocaris vandoverae from the Mariana Back Arc Basin and Chorocaris paulexa from the East Pacific Rise	eura of fourth and fifth abdominal somites not serrated
Rimicaris hybisa	Rostrum short but conspicuous; dorsal organ with a pore; antepenultimate segment hird maxilliped with 1–2 distal movable spines
R. exoculata-R. kairei complex (R. exoculata in the Atlantic, F.	ostrum absent; dorsal organ without pores; antepenultimate segment of third

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Author Contributions

Analyzed the data: AV DK AL. Wrote the paper: AV DK.

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